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Molecular evolution of *HoxA13* and the multiple origins of limbless morphologies in amphibians and reptiles

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Abstract

Developmental processes and their results, morphological characters, are inherited through transmission of genes regulating development. While there is ample evidence that cis-regulatory elements tend to be modular, with sequence segments dedicated to different roles, the situation for proteins is less clear, being particularly complex for transcription factors with multiple functions. Some motifs mediating protein-protein interactions may be exclusive to particular developmental roles, but it is also possible that motifs are mostly shared among different processes. Here we focus on *HoxA13*, a protein essential for limb development. We asked whether the *HoxA13* amino acid sequence evolved similarly in three limbless clades: Gymnophiona, Amphisbaenia and Serpentes. We explored variation in ω (dN/dS) using a maximum-likelihood framework and *HoxA13* sequences from 47 species. Comparisons of evolutionary models provided low ω global values and no evidence that *HoxA13* experienced relaxed selection in limbless clades. Branch-site models failed to detect evidence for positive selection acting on any site along branches of Amphisbaenia and Gymnophiona, while three sites were identified in Serpentes. Examination of alignments did not reveal consistent sequence differences between limbed and limbless species. We conclude that *HoxA13* has no modules exclusive to limb development, which may be explained by its involvement in multiple developmental processes.

Keywords: development, evolution, *HoxA13*, molecular signatures, limblessness.

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Introduction

Evolution of morphological diversity has fascinated biologists, but only in the past half-century the investigation of mechanisms underlying the origin and establishment of specific phenotypes became possible through the combination of genetics, evolution and developmental biology in the field so-called Evo-Devo (*Evolution of Development*, see Raff, 2000; Hall, 2012). Consolidation of Evo-Devo fostered the search for variations in develop-

mental processes likely responsible for the distribution of heritable phenotypic variance that potentially could be molded by natural selection (Brakefield, 2006; 2011).

Differences in gene expression during developmental processes often can be explained by sequence variation in cis-regulatory elements but coding region variations of transcription factors are also a possible source of developmental variation. Mutations in cis-regulatory elements have been argued to be a more likely source of adaptive genetic variation, a trend that is often found when investigating cis-regulatory evolution for phenotypic divergence (Stern, 2000; Wray, 2007; Mansfield, 2013). Recent studies, however, provide a wide range of evidence supporting

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the contribution of mutations in coding regions of transcription factor genes for the diversification of phenotypes (Galant and Carroll, 2002; Lynch *et al.*, 2008; Crow *et al.*, 2009; Brayer *et al.*, 2011). Such findings imply that transcription factors do not remain functionally equivalent during evolution (Galant and Carroll, 2002; Ronshaugen *et al.*, 2002; Lynch *et al.*, 2008; Crow *et al.*, 2009), and that the adaptive evolution of transcription factors proteins may be involved in the origin of new phenotypes (Lynch *et al.*, 2004; Lynch and Wagner, 2008; Crow *et al.*, 2009; Brayer *et al.*, 2011). A disparity emerging from current literature is that there is ample evidence that cis-regulatory elements tend to be modular, with dedicated sequence segments for different developmental roles, but the role of transcription factor proteins is more complex due to the potential for pleiotropic constraints. Discussions about sequence modules dedicated to specific developmental roles in transcription factor proteins is particularly complex for homeotic proteins involved in multiple functions, and two alternative scenarios emerge (Sivanantharajah and Percival-Smith, 2015): there may be specific motifs mediating protein-protein interactions that are exclusive to particular developmental roles, while other motifs within the same protein are shared among different developmental processes, for instance the highly conserved homeodomain. In the second scenario, changes in a given sequence segment that plays two developmental roles likely affect both processes, as well as their results (*i.e.* the morphological characters established in the developing embryo), so that any motif involved in multiple developmental processes would be expected to be under pleiotropic constraint (for recent discussions about the topic see Pavlicev and Wagner, 2012; Pavlicev and Widder, 2015).

The counterpoint between the presence of motifs dedicated to specific developmental roles and a pleiotropic constraint that may be imposed by the commitment of some motifs to multiple functions may be investigated through combination of two approaches: 1) a transcription factor protein known to be involved in more than one developmental process, and 2) a phylogenetic framework where one developmental process was suppressed but another remains effective in the organism. HOX proteins emerge as good candidates because they play central roles during embryo development in the specification of structures along the vertebrate anterior-posterior body axis, and have also acquired several functions including development of limbs and the urogenital system (Hsieh-Li *et al.*, 1995; Taylor *et al.*, 1997; Kobayashi and Behringer, 2003; Sivanantharajah and Percival-Smith, 2015). The transcription factor *HoxA13*, in particular, is essential for several functions (Fromental-Ramain *et al.*, 1996; Innis *et al.*, 2002; Shou *et al.*, 2013): during limb development it regulates formation of digits, phalangeal joints and carpal/tarsal elements (Stadler *et al.*, 2001; Knosp *et al.*, 2004, 2007; Perez *et al.*, 2010); during organogenesis, it modulates development of

digestive and urogenital tracts, including differentiation of the mammalian female reproductive system (Taylor *et al.*, 1997). Moreover, there is evidence for pleiotropy effects on *HoxA13* and mutations in this gene simultaneously affect the development of the urogenital system and the limbs (Mortlock and Innis, 1997; Goodman *et al.*, 2000). Specifically, in mice a 50 base-pair deletion at the first exon of *HoxA13* results in hypodactyly (Mortlock *et al.*, 1996), while the expansion of an N-terminal polyalanine in this first exon is associated with limb and genitourinary abnormalities in humans (Goodman *et al.*, 2000).

A good evolutionary scenario to test for the presence of sequence segments exclusively dedicated to a given function in the pleiotropic *HoxA13* gene is the recurrent evolution of limbless morphologies in basal lineages of Tetrapoda. Evolution of snakelike morphologies in Lepidosauria and Lissamphibia is characterized by body elongation and limb loss (Woltering, 2012). In this scenario, the presence and identity of limb-specific sequence segments in the protein would be supported by identification of sequence changes in *HoxA13* that are common to different limbless lineages. In contrast, the likelihood that all motifs involved in limb development are also committed to other developmental functions would receive support if it is found that there are no consistent sequence differences between limbed and limbless species. In the present study we compared the molecular sequence variation of *HoxA13* among three tetrapod snakelike lineages that independently lost limbs: Serpentes, Amphisbaenia and Gymnophiona. We used both likelihood-ratio tests implemented in PAML (Yang, 2007) and visual inspections of the alignments to compare *HoxA13* sequences among these three snakelike lineages.

Material and Methods

Tissue samples were obtained from Museum Herpetological Collections or Private Zoological Collections, and the molecular database for *HoxA13* assembled was complemented with sequences available at GenBank (see Table S1). Genomic DNA was extracted from the tissue samples using the DNeasy Tissue Kit (Qiagen), following the manufacturer's instructions. In this study we focused on the first exon of *HoxA13* due to described phenotypic effects of mutations in this region and the general conservation of the homeodomain which is coded for in exon 2 (Mortlock *et al.*, 1996; Goodman *et al.*, 2000). Gene fragments of *HoxA13* exon-1 (375 to 455 bp), were amplified from one individual of each species using the following conditions: 50 to 100 ng of DNA and primers at 10 μ M concentration combined with PCR Master Mix (Reddymix, 2.5 mM MgCl₂; Abgene, Inc.) to a final reaction volume of 50 μ L; thermocycling conditions consisted of 30 or 35 cycles (1 min at 94 °C, 1 min of annealing at 48–52 °C, 1 min at 72 °C), followed by a terminal extension step (5 to 8 min at 72 °C). Primers used to amplify

HoxA13 were synthesized based on sequences from Mortlock *et al.* (2000), as follows:

F1, 5-CTATGACAGCCTCCGTGCTC-3;

F2, 5-ATCGAGCCCACCGTCATGTTTCTCTACGAC-3;

R1, 5-CGAGCTCTGTGCCGTCGCCGAGTAGGGACT-3;

R2, 5-TGGTAGAAAGCAAACCTCCTTG-3.

The PCR products were gel-purified using the Wizard SV Gel and PCR Clean-up System (Promega) according to the manufacturer's instructions. Genes were cloned using the pGEM-T vector system (Promega) and *E. coli* -competent cells (DH5 α). Three to eight clones of each species were sequenced to control for errors during PCR amplification. Sequencing was performed in both directions with the vector primers T7 and SP6 (sequence according to technical manual pGEM-T Vector System-Promega), using an ABI 3730 DNA Analyzer (Applied Biosystems).

In total, we sequenced *HoxA13* from 23 species of Squamata and Lissamphibia, and downloaded 24 additional *HoxA13* sequences from GenBank. All sequences obtained were aligned using ClustalW algorithm (Thompson *et al.*, 1994) implemented in the software BioEdit sequence alignment editor, and the alignment was manually improved based on amino acid translated sequences. The alignment was first visually inspected for indels, and no conspicuous patterns common to all three snakelike lineages were identified. In order to investigate natural selection acting on the sequence of *HoxA13* exon-1 in the three snakelike lineages that represent independent limb losses, we performed molecular evolution analyses in three separate data sets (Figure 1): A) limbed lizards *versus* Amphisbaenia; B) limbed lizards *versus* Serpentes; C) limbed amphibians (anurans and salamanders) *versus* Gymnophiona (caecilians). Such analyses are implemented

under a phylogenetic framework, so we adopted the phylogenetic hypothesis proposed by Pyron *et al.* (2013) for Squamata and that proposed by Pyron and Wiens (2011) for Lissamphibia.

In order to investigate the possible roles of relaxed or directional selections on the evolution of *HoxA13* in the limbless lineages, we explored the variation in ω , the ratio of the rate of non-synonymous substitutions (dN) to the rate of synonymous substitutions (dS), in a maximum likelihood framework using the codeml program implemented in PAML (Yang, 2007). In these tests, indels were deleted, so that only portions of the alignment that were unambiguous were used for the dN/dS analyses. We conducted the “free ratio” branch model (Model 1), which assumes an independent ω ratio for each branch. Although very effective, this test is generally conservative once this approach averages substitution rates over all amino acid sites in the sequence (Bamshad and Wooding, 2003). As most amino acid sites are expected to be highly conserved and adaptive evolution most likely affects only a few sites, we also applied the branch-site model that estimates rates of evolution in a codon-by-codon basis on a specific branch of the tree. In addition, with this model we could test whether the limbless lineages share common sites under positive selection that could have evolved convergently. In that sense, models of molecular signatures representing directional selection or neutral evolution were tested in the *HoxA13* of snakes, amphisbaenians and caecilians using the branch-site model A implemented in PAML (Zhang *et al.*, 2005; Yang, 2007). This model tests for positive selection on individual sites along a specific lineage of the tree, called foreground branch, where the other lineages are background branches. In our case, the limbless lineages were labeled as fore-

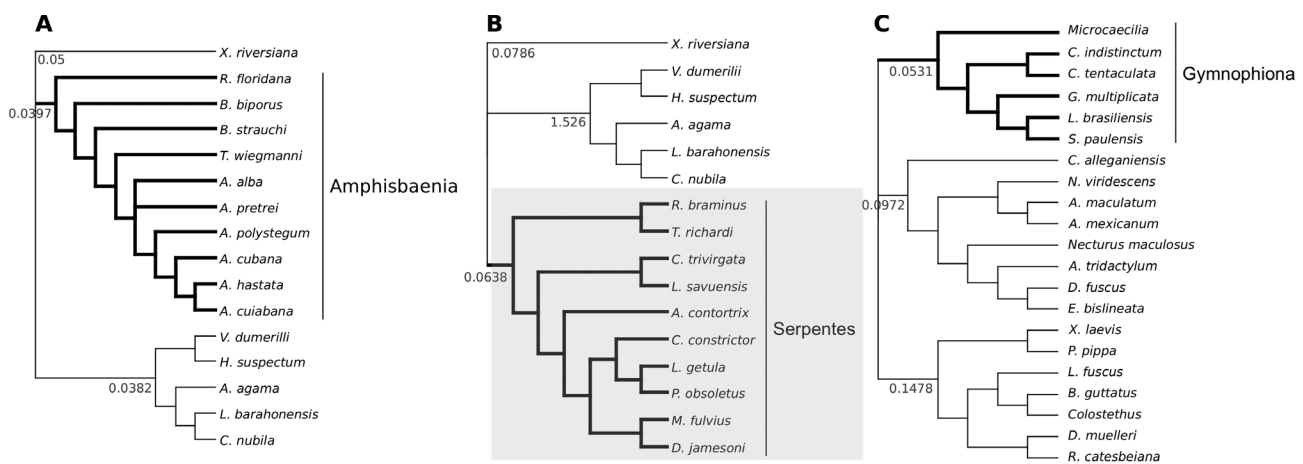


Figure 1 - Topology used for test models of molecular signatures of *HoxA13* in limbless lineages. The models of molecular signatures of *HoxA13* were tested in the three main comparisons. A) limbed lizards *versus* Amphisbaenia; B) limbed lizards *versus* Serpentes; C) limbed amphibians (anurans and salamanders) *versus* Gymnophiona. The tree to conduct the analyses of variable ω among lineages and sites is based on published literature (Pyron *et al.*, 2013 for Squamata and Pyron and Wiens, 2011 for Lissamphibia). The bold branches correspond to the groups labeled as ‘foreground branches’. The shaded tree represents the one where the branch-site model identified signatures of positive selection. The values represented in each branch correspond an independent ω ratio that corresponds to ratio of the rate of non-synonymous substitutions (dN) to the rate of synonymous substitutions (dS) in a maximum likelihood framework.

ground branches as depicted on Figure 1. In this method, codon sites are categorized into four classes 0, 1, 2a, and 2b with respective proportions of p_0 , p_1 , p_{2a} , and p_{2b} . In site class 0, negative selection is assumed on both the foreground and background branches, with $0 < \omega_0 < 1$. In site class 1, codons are assumed to evolve neutrally in all lineages, with $\omega_1 = 1$. In class 2a, it is assumed that positive selection operates on the foreground branch with $\omega_2 > 1$, and that negative selection operates on the background branches, with $\omega = \omega_0$. Finally in class 2b, positive selection is allowed on the foreground branch with $\omega_2 > 1$, but no selection is assumed for the background branches. The corresponding null model is the same as model A, except that no selection is assumed on the foreground branch in classes 2a and 2b, and ω_2 is fixed to 1. The nested models were compared using the likelihood-ratio test (LRT), and the level of significance was 0.05. If the null hypothesis is rejected, a Bayes empirical approach is used to calculate the posterior probabilities that each site has evolved under positive selection on the foreground lineage (Yang *et al.*, 2005). Each branch-site model was run multiple times, with three starting ω values (0.5, 1, and 2) to check the existence of multiple local optima, as recommended.

Because the branch-site model implemented in PAML can be limiting due to the necessary specification of foreground lineages and the assumption that $\omega = 1$ for all background lineages (Zhang *et al.*, 2005), we also examined *HoxA13* for signatures of episodic selection using the mixed model of evolution (MEME, Murrell *et al.*, 2012) and the fixed-effect likelihood (FEL) model of molecular evolution performed with HyPhy in Datamonkey server (Delpont *et al.*, 2010). The FEL model estimates the ratio of dN/dS on a site-by-site basis, without assuming an a priori distribution across sites. The MEME model allows the distribution of the estimated ω value to vary among sites and branches, and identifies episodes of positive selection that affect only a subset of lineages.

Results

We sequenced the exon-1 of *HoxA13* in a total of 23 species, being five species of Amphisbaenia, seven of

Caudata, five of Gymnophiona, and six of Anura (see details in Supplementary Table S1). The database was complemented with 24 published sequences of *HoxA13* downloaded from GenBank: five species of Amphisbaenia and six of limbed lizards, ten snake species, one species of Caudata, one of Gymnophiona and one of Anura (details given in Supplementary Table S1). Analyses of *HoxA13* molecular evolution were implemented as follows: (1) limbed lizards *versus* Amphisbaenia (Figure 1A); (2) limbed lizards *versus* Serpentes (Figure 1B); and (3) limbed amphibians *versus* Gymnophiona (Figure 1C). To address the global evolutionary pressure acting on the *HoxA13* gene in these lineages we obtained their ω (dN/dS) throughout the model M1 (“free model”) in the branch model. The ω values obtained for limbless lineages are much lower than the neutral rate and ranged from 0.0397 to 0.0638 (Figure 1), being equal or lower than the corresponding values of the other lineages (0.0382 to 1.526). These values indicate that *HoxA13* in the limbless lineages experienced strong purifying selection, even when “released” from one important function (limb and digits formation).

To investigate whether there are sites under positive selection and to determine whether some amino acid sites could have undergone convergent contributions to developmental changes in the limbless lineages, we applied the robust branch-site model. The likelihood-ratio tests (LRTs) support the model of adaptive evolution in *HoxA13* only in Serpentes, while in Amphisbaenia and in Gymnophiona the fit of the null model for neutral evolution was not significantly different than the alternative model (Table 1). The evidence for positive selection in *HoxA13* detected by the branch-site model indicates that, when the stem lineage of Serpentes was labeled as foreground branch, the model estimating a class of sites with a ω value greater than 1 (model A) had a significantly better fit than the null model (Table 1). In this model, three codons were identified as being under positive selection in *HoxA13* only in the Serpentes lineage: sites 46, 88 and 121 (Table 1). Models of non-neutral evolution were not supported in amphisbaenians and caecilians, as the likelihoods of a branch-site model and the null one were not statistically different in these lineages

Table 1 - Summary of likelihood-ratio tests performed using branch-site models implemented in PALM. Log-likelihood values of different models tested using Amphisbaenia, Serpentes or Gymnophiona labeled as foreground branches. LnL corresponds to the likelihood value. Sites inferred under positive selection in Serpentes had posterior probabilities values of 0.64, 0.76 and 0.51, respectively.

Foreground branch	Model	LnL	LRT	p	Positively selected sites
Serpentes	Model <u>A</u>	-1620.1549	3.967	0.046	46, 88, 121
	Null	-1622.1386			
Amphisbaenia	Model <u>A</u>	- 1287.5078	0	> 0.05	-
	Null	- 1287.5078			
Gymnophiona	Model <u>A</u>	-1777.4047	0	> 0.05	-
	Null	-1777.4047			

(Table 1), indicating that limb loss did not imprint consistent sequence differences onto the *HoxA13* gene between limbed and limbless lineages.

The absence of consistent sequence differences in *HoxA13* between limbed and limbless lineages was also corroborated in the analyses performed using MEME, where we identified episodic positive selection only in codon 66 in the ancestral lineage of all snakes, and in two amphisbaenian species (*A. polystegum* and *A. cuiabana*), as shown in the Supplementary Figure S1. This codon was not the same identified in Serpentes by the branch-site model implemented in PAML; the FEL model did not identify any codon under positive selection in our dataset.

Discussion

This study investigated whether there are limb-specific sequence elements in the transcription factor protein *HoxA13*. We used as model system the recurrent independent evolution of limbless lineages within Tetrapoda. In this framework, identification of molecular signatures in *HoxA13* that are common to independently derived limbless lineages would suggest the presence of limb-specific sequence segments in the protein, while no consistent sequence differences between limbed and limbless species suggests that all motifs involved in limb development are also committed to other developmental functions. The conceptual basis underpinning the first prediction relies on evidence that genetic elements dedicated to the development of a particular structure tend to get lost when the corresponding structure is lost in evolution (Graur and Li, 1999), as observed in the loss of pelvic spines in lake morphs of sticklebacks (Bell, 1987) and of penile spines in humans (Reno *et al.*, 2013). In these examples, a dedicated cis-regulatory element has been lost in evolution coincidentally with the loss of the morphological structure, suggesting the existence of sequence modules dedicated to specific developmental processes. We apply this reasoning to coding regions and investigated amino acid sequence variation in the first exon of *HoxA13* to test for specific amino acid sequence motifs in transcription factor proteins that may be exclusively dedicated to certain biological roles.

Analyses of molecular evolution in the pleiotropic *HoxA13* gene were applied to the evolutionary scenario of recurrent evolution of limbless tetrapod lineages, here represented by three snakelike clades (see Woltering, 2012): Gymnophiona (Lissamphibia), Amphisbaenia and Serpentes (Lepidosauria). We investigated the overall constraint, the site-specific evolutionary rates, and evidence of positive selection acting on *HoxA13*. None of our analyses revealed a signal specific to and shared by the three snake-like clades investigated. Specifically, our comparisons of evolutionary models based on likelihood-ratio tests provided low global values of ω , and the branch-site model failed to detect evidence of positive selection acting on any

site along the branch leading to the Amphisbaenia and the Gymnophiona lineages, identifying three sites evolving under positive selection of *HoxA13* only in snakes. From these results we conclude that the first exon of *HoxA13* does not have limb-specific sequence motifs, and propose that all protein-protein interaction motifs of the *HoxA13* protein necessary for limb development are also involved in the establishment of other structures. Nonetheless, we recognize that *HoxA13* limb-specific motifs may actually exist although remaining unidentified under the approach used here, though unlikely given the sequence evidence presented in this study. There could still be limb-specific motifs in the C-terminal tail that is coded for by exon 2 and which is not covered by the data analyzed here.

Evolution of snakelike tetrapods involves interlocked changes in different traits: limb reduction/loss occurs simultaneously with body elongation (Gans, 1975; Lande, 1978). Two major clades represented here, Lepidosauria and Lissamphibia, differ in the evolutionary patterns of such morphological transitions. Lissamphibia has fewer independent events of snakelike evolution (three transitions in salamanders [Parra-Olea and Wake, 2011] plus the origin of Gymnophiona [Pyrn and Wiens, 2011; San Mauro *et al.*, 2014]) when compared to Lepidosauria (at least 26 independent origins; Wiens *et al.*, 2006). These transitions likely involved changes in genes underlying the development of the modified structures. For example, snakelike organisms exhibit an acceleration of the somitogenesis clock rhythm, a delay in the shrinkage of pre-somitic mesoderm (PSM), changes in expression domains of *Hox* genes, and a differential interpretation of *Hox* codes by downstream genes in the pre-caudal region (Cohn and Tickle, 1999; Woltering *et al.*, 2009; Di-Poi *et al.*, 2010; Woltering, 2012). These modifications in developmental processes culminate on increased numbers of vertebrae and the consequent body elongation, as well as a vertebral deregionalization associated with limb loss (Woltering, 2012; but see Head and Polly, 2015). Variation in expression domains of *Hox* genes is particularly relevant in this context because *Hox* proteins are essential for morphogenesis and patterning of the vertebrate skeleton during embryo development (Krumlauf, 1994; Pearson *et al.*, 2005). The gene *HoxA13* in particular is expressed in both the developing autopodia, genital tubercle mesenchyme, and the hindgut and cloacal region, the later resulting in the development of the intestine and urogenital tracts from its terminal end (Dollé *et al.*, 1991a,b; Warot *et al.*, 1997). For our study this gene is an ideal candidate not only because it is involved in the development of traits remarkably modified in snakelike morphologies (*e.g.* limbs, digestive system and reproductive tract), but also because changes in *HoxA13* likely encompass pleiotropy due to its involvement in different developmental processes (for pleiotropic effects of mutations in *Hox* see Mortlock and Innis, 1997).

The absence of consistent differences between limbed and limbless species in sequence elements of the transcription factor *HoxA13* favors the scenario where limb-development motifs in the protein are also committed to other developmental processes. Evidence for this interpretation is also provided by mutations of *HoxA13* that are causal for the so-called hand-foot-genital syndrome (Mortlock and Innis, 1997). Such pleiotropic effect of a limb gene on penile structures is explained by the likely serial homology of hind limbs and external genitalia in squamates (Tschopp *et al.*, 2014). Such homology reinforces that changes in limb-related amino acids of the *HoxA13* protein are likely to also affect the development of the penis, implying associated fitness costs.

The genetic and evolutionary connections between external genitalia (penis) and limbs may explain the absence of limb-specific variation in the *HoxA13* amino acid sequence for at least for the two lepidosaurian clades, Serpentes and Amphisbaenia, which have limb-related penises. At this point we do not have a plausible explanation for the evidence for directional selection on the *HoxA13* coding sequence in the stem lineage of snakes. It is important to note, though, that the function of *Hox* genes has undergone substantial reorganization in squamates and snakes in particular (Di-Poï *et al.*, 2010), so there might be other structures in snakes where *HoxA13* has acquired novel or modified functions. For example, a characteristic of the Serpentes lineage is the presence cloacal scent glands, a morphological trait evoked previously as a possible candidate for a new function that co-opted *HoxA13* in the clade (Kohlsdorf *et al.*, 2008). The hypothesis that *HoxA13* may be involved in the development of snake-specific cloacal scent glands has been tested by our group, but no *HoxA13* transcripts expressed in these rudiments have been identified using RT-PCR (results not shown). The functional significance of the positive selection detected in the *HoxA13* of Serpentes remains therefore under investigation.

The explanation for the lack of a limb-loss signal in the *HoxA13* amino acid sequence based on the genital-limb connection described by Tschopp *et al.* (2014) is not applicable to caecilians. These animals do universally have a male copulatory organ, the phalloidium, but this structure is an inverted cloaca and not a body appendage (Gower and Wilkinson, 2002). Given that there are no obvious limb-related appendages in caecilians, one could expect a signal of limb loss in these animals. In fact, there is a substitution from isoleucine (I) to a methionine (M) at position 6 and a deletion of three amino acids (two alanines and a glutamine - AAQ) at positions 76 to 78 of the alignment that are specific to caecilians. However, it is hard to dissect functional significance from phylogenetic signal in these patterns because our dataset lacks additional snakelike and penis-less amphibians other than caecilians, a morphology represented by limbless urodeles.

In summary, we conclude that there is no evidence for limb-specific/exclusive sequence motifs in the *HoxA13* amino acid sequence coded for by exon 1, at least in squamates, so that all sequence elements of this part of the protein that are necessary for limb development seem also committed to other developmental processes. We note though, that the C-terminal tail that is coded for by exon 2 is not covered by this study and, thus, we cannot exclude the presence of limb-specific motifs in this part of the *HoxA13* molecule.

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References

- Bamshad M and Wooding SP (2003) Signatures of natural selection in the human genome. *Nat Rev Genet* 4:99-111.
- Bell MA (1987) Interacting evolutionary constraints in pelvic reduction of threespine sticklebacks, *Gasterosteus aculeatus* (Pisces, Gasterosteidae). *Biol J Linn Soc* 31:347-382.
- Brayer KJ, Lynch VJ and Wagner GP (2011) Evolution of a derived protein-protein interaction between HoxA11 and Foxo1a in mammals caused by changes in intramolecular regulation. *Proc Natl Acad Sci U S A* 108:E414-E420.
- Brakefield PM (2006) Evo-devo and constraints on selection. *Trends Ecol Evol* 21:362-368.
- Brakefield PM (2011) Evo-devo and accounting for Darwin's endless forms. *Phil Trans R Soc B Biol Sci* 366:2069-2075.
- Cohn M and Tickle C (1999) Developmental basis of limblessness and axial patterning in snakes. *Nature* 399:474-479.
- Crow KD, Amemiya CT, Roth J and Wagner GP (2009). Hypermutability of HoxA13a and functional divergence from its paralog are associated with the origin of a novel developmental feature in zebrafish and related taxa (cypriniformes). *Evolution* 63:1574-1592.
- Delpont W, Poon AFY, Frost SDW and Kosakovsky Pond SL (2010) Datamonkey 2010: A suite of phylogenetic analysis tools for evolutionary biology. *Bioinformatics* 26:2455-2457.
- Di-Poï N, Montoya-Burgos JI, Miller H, Pourquié O, Milinkovitch MC and Duboule D (2010) Changes in Hox genes' structure and function during the evolution of the squamate body plan. *Nature* 464:99-103.
- Dollé P, Izpisua-Belmonte JC, Brown JM, Tickle C and Duboule D (1991a) HOX-4 genes and the morphogenesis of mammalian genitalia. *Genes Dev* 5:1767-1776.

- Dollé P, Izpisua-Belmonte JC, Boncinelli E and Duboule D (1991b) The Hox-4.8 gene is localized at the 5' extremity of the Hox-4 complex and is expressed in the most posterior parts of the body during development. *Mech Dev* 36:3-13.
- Fromental-Ramain C, Warot X, Messadecq N, LeMeur M, Dollé P and Chambon P (1996) Hoxa-13 and Hoxd-13 play a crucial role in the patterning of the limb autopod. *Development* 122:2997-3011.
- Galant R and Carroll SB (2002) Evolution of a transcriptional repression domain in an insect Hox protein. *Nature* 415:910-913.
- Gans C (1975) Tetrapod limblessness: Evolution and functional corollaries. *Am Zool* 15:455-467.
- Goodman FR, Bacchelli C, Brady AF, Brueton LA, Fryns JP, Mortlock DP and Scambler PJ (2000) Novel *HOXA13* mutations and the phenotypic spectrum of Hand-Foot-Genital Syndrome. *Am J Hum Genet* 67:197-202.
- Gower DJ and Wilkinson M (2002) Phallus morphology in caecilians (Amphibia, Gymnophiona) and its systematic utility. *Bull Nat Hist Mus Zool* 68:143-154.
- Graur D and Li WH (1999) Fundamentals of Molecular Evolution. 2nd edition. Sinauer Associates Inc., Sunderland.
- Hall BK (2012) Evolutionary developmental biology (Evo-Devo): Past, present, and future. *Evolution: Educ Outreach* 2:184-193.
- Head JJ and Polly PD (2015) Evolution of the snake body form reveals homoplasy in amniote Hox gene function. *Nature* 520:86-89.
- Hsieh-Li HM, Witte DP, Weinstein M, Branford W, Li H, Small K and Potter SS (1995) Hoxa 11 structure, extensive antisense transcription, and function in male and female fertility. *Development* 121:1373-1385.
- Innis JW, Goodman FR, Bacchelli C, Williams TM, Mortlock DP, Sateesh P and Gutmacher AE (2002) A *HOXA13* allele with a missense mutation in the homeobox and a dinucleotide deletion in the promoter underlies Gutmacher syndrome. *Hum Mutat* 19:573-574.
- Knosp WM, Scott V, Bächinger HP and Stadler HS (2004) *HOXA13* regulates the expression of bone morphogenetic proteins 2 and 7 to control distal limb morphogenesis. *Development* 131:4581-4592.
- Knosp WM, Saneyoshi C, Shou S, Bächinger HP and Stadler HS (2007) Elucidation, quantitative refinement, and *in vivo* utilization of the *HOXA13* DNA binding site. *J Biol Chem* 282:6843-6853.
- Kobayashi A and Behringer RR (2003) Developmental genetics of the female reproductive tract in mammals. *Nat Rev Genet* 4:969-980.
- Kohlsdorf T, Cummings MP, Lynch VJ, Stopper GF, Takahashi K and Wagner GP (2008) A molecular footprint of limb loss: Sequence variation of the autopodial identity gene Hoxa-13. *J Mol Evol* 67:581-593.
- Krumlauf R (1994) Hox genes in vertebrate development. *Cell* 78:191-201.
- Lande R (1978) Evolutionary mechanism of limb loss in tetrapods. *Evolution* 32:73-92.
- Lynch VJ and Wagner GP (2008) Resurrecting the role of transcription factor change in developmental evolution. *Evolution* 62:2131-2154.
- Lynch VJ, Roth JJ, Takahashi K, Dunn CW, Nonaka DF, Stopper GF and Wagner GP (2004) Adaptive evolution of HoxA-11 and HoxA-13 at the origin of the uterus in mammals. *Proc R Soc Lond B Biol Sci* 271:2201-2207.
- Mansfield JH (2013) Cis-regulatory change associated with snake body plan evolution. *Proc Natl Acad Sci USA* 110:10473-10474.
- Mortlock DP and Innis JW (1997) Mutation of *HOXA13* in hand-foot-genital syndrome. *Nat Genet* 15:179-180.
- Mortlock DP, Post LC and Innis JW (1996) The molecular basis of hypodactyly (Hd): A deletion in *Hoxa13* leads to arrest of digital arch formation. *Nat Genet* 13:284-289.
- Mortlock DP, Sateesh P and Innis JW (2000) Evolution of N-terminal sequences of the vertebrate Hoxa-13 protein. *Mamm Genome* 11:151-158.
- Murrell B, Wertheim JO, Moola S, Weighill T, Scheffler K and Kosakovsky SL (2012) Detecting diversifying sites subject to episodic diversifying selection. *PLoS Genet* 8:e1002764.
- Parra-Olea G and Wake DB (2001) Extreme morphological and ecological homoplasy in tropical salamanders. *Proc Natl Acad Sci U S A* 98:7888-7891.
- Pavlicev M and Wagner GP (2012) A model of developmental evolution: Selection, pleiotropy and compensation. *Trends Ecol Evol* 27:316-322.
- Pavlicev M and Widder S (2015) Wiring for independence: Positive feedback motifs facilitate individuation of traits and development and evolution. *J Exp Zool Mol Dev Evol* 324B:104-113.
- Pearson JC, Lemons D and McGinnis W (2005) Modulating Hox gene functions during animal body patterning. *Nat Rev Genet* 6:893-904.
- Perez WD, Weller CR, Shou S and Stadler HS (2010) Survival of *hoxa13* homozygous mutants reveals a novel role in digit patterning and appendicular skeletal development. *Dev Dyn* 239:446-457.
- Pyron AR and Wiens JJ (2011) A large-scale phylogeny of Amphibia including over 2800 species, and a revised classification of extant frogs, salamanders, and caecilians. *Mol Phylogenet Evol* 61:543-583.
- Pyron RA, Burbrink FT and Wiens JJ (2013) A phylogeny and revised classification of Squamata, including 4161 species of lizards and snakes. *BMC Evol Biol* 13:e93.
- Raff RA (2000) Evo-devo: The evolution of a new discipline. *Nat Rev Genet* 1:74-79.
- Reno PL, McLean CY, Hines JE, Capellini TD, Bejerano G and Kingsley DM (2013) A penile spine/vibrissa enhancer sequence is missing in modern and extinct humans but is retained in multiple primates with penile spines and sensory vibrissae. *PLoS One* 8:e84258.
- Ronshaugen M, McGinnis N and McGinnis W (2002) Hox protein mutation and macroevolution of the insect body plan. *Nature* 415:914-917.
- San Mauro D, Gower DJ, Müller H, Loader SP, Zardoya R, Nussbaum RA and Wilkinson M (2014) Life-history evolution and mitogenomic phylogeny of caecilian amphibians. *Mol Phylogenet Evol* 73:177-189.
- Shou S, Carlson HL, Perez WD and Stadler HS (2013) *HOXA13* regulates *Aldh1a2* expression in the autopod to facilitate interdigital programmed cell death. *Dev Dyn* 242:687-698.
- Sivanantharajah L and Percival-Smith A (2015) Differential pleiotropy and HOX functional organization. *Dev Biol* 398:1-10.

- Stadler HS, Higgins KM and Capecchi MR (2001) Loss of Eph-receptor expression correlates with loss of cell adhesion and chondrogenic capacity in Hoxa13 mutant limbs. *Development* 128:4177-4188.
- Stern DL (2000) Perspective: Evolutionary developmental biology and the problem of variation. *Evolution* 54:1079-1091.
- Taylor HS, Heuvel GV and Igarashi P (1997) A conserved Hox axis in the mouse and human female reproductive system: Late establishment and persistent adult expression of the Hoxa cluster genes. *Biol Reprod* 57:1338-1345.
- Thompson JD, Higgins DG and Gibson TJ (1994) CLUSTAL W: Improving the sensitivity of progressive multiple sequence alignment through sequence weighting, position-specific gap penalties and weight matrix choice. *Nucleic Acids Res* 22:4673-4680.
- Tschopp P, Sherratt E, Sanger TJ, Groner AC, Aspiras AC, Hu JK and Tabin CJ (2014) A relative shift in cloacal location repositions external genitalia in amniote evolution. *Nature* 516:391-394.
- Warot X, Fromental-Ramain C, Fraulob V, Chambon P and Dollé P (1997) Gene dosage-dependent effects of the Hoxa-13 and Hoxd-13 mutations on morphogenesis of the terminal parts of the digestive and urogenital tracts. *Development* 124:4781-479.
- Wiens JJ, Brandley MC and Reeder TW (2006) Why does a trait evolve multiple times within a clade? Repeated evolution of snakelike body form in squamate reptiles. *Evolution* 60:93-111.
- Woltering JM (2012) From lizard to snake; behind the evolution of an extreme body plan. *Curr Genomics* 13:e289.
- Woltering JM, Vonk FJ, Müller H, Bardine N, Tudeau IL, de Bakker MA and Richardson MK (2009) Axial patterning in snakes and caecilians: Evidence for an alternative interpretation of the Hox code. *Dev Biol* 332:82-89.
- Wray GA (2007) The evolutionary significance of cis-regulatory mutations. *Nature Rev Genet* 8:206-216.
- Yang Z, Wong WS and Nielsen R (2005) Bayes empirical Bayes inference of amino acid sites under positive selection. *Mol Biol Evol* 22:1107-1118.
- Yang Z (2007) PAML 4: Phylogenetic analysis by maximum likelihood. *Mol Biol Evol* 24:1586-1591.
- Zhang J, Nielsen R and Yang Z (2005) Evaluation of an improved branch-site likelihood method for detecting positive selection at the molecular level. *Mol Biol Evol* 22:2472-2479.

Supplementary Material

The following online material is available for this article:
Table S1- List of species used in the bioinformatic analyses.

Figure S1 - Results from HyPhy analyses in the Data-monkey server.

This material is available as part of the online article from <http://www.scielo.br/gmb>.

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